



3.3.2. NUMBER OF BOOKS AND CHAPTERS IN EDITED VOLUMES/BOOKS PUBLISHED AND PAPERS PUBLISHED IN NATIONAL/ INTERNATIONAL CONFERENCE PROCEEDINGS PER TEACHER DURING THE YEAR

NAMES OF THE AUTHORS

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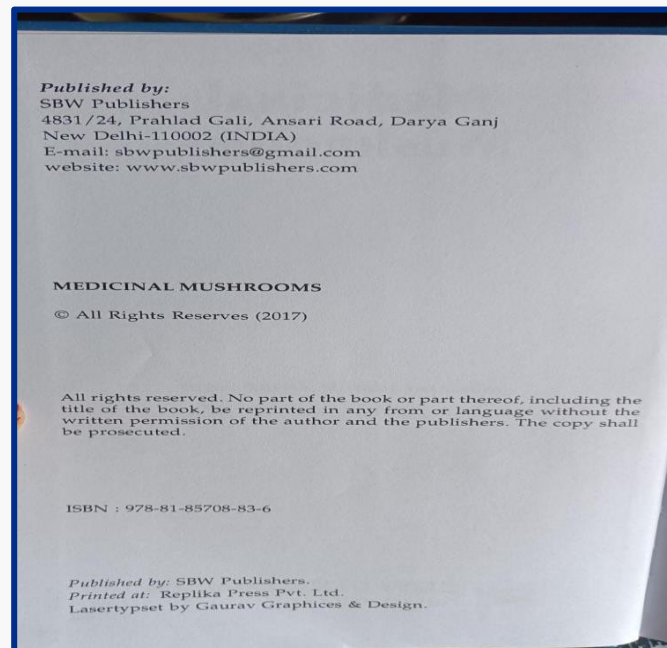
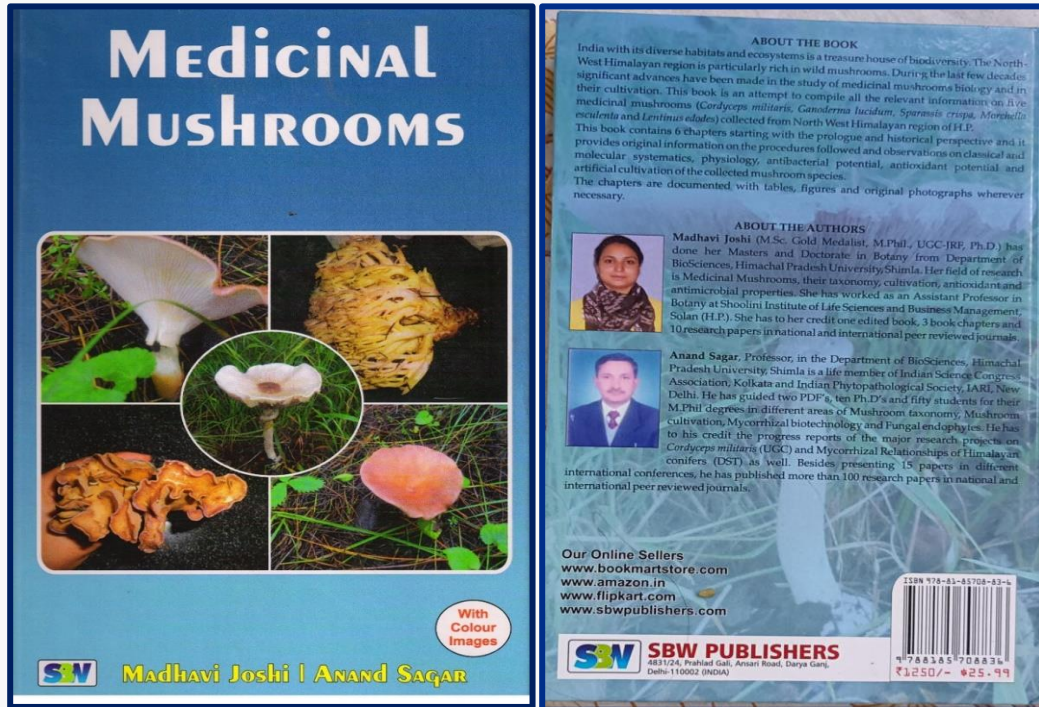
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- Dr. Madhavi Joshi- Department of Botany
Title of Book: Medicinal Mushrooms
Year: 2017-18
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Medicinal Mushrooms

3.3.2 Amplification of genomic DNA

Amplification of genomic DNA was performed by polymerase chain reaction (PCR). Amplification of rRNA gene for assessing ITS length variation was done using universal primers ITS1 (TCCGTAGGTGAACCTGCGG) and ITS4 (TCCCGCTTATTGAT-ATGC) as described by White *et al.* (1990).

The PCR amplification was carried out in 0.2 ml PCR tubes. The reaction mixture (50 µl total volume) of PCR contained Taq buffer 0.5 µl, dNTP 0.4 µl, primer1 and primer4 1.5 µl, Taq DNA Polymerase 1.5 units, 2.5 µl template DNA and 37.6 µl double distilled water. The tubes were then placed on thermocycler model C1000 (BIORAD, Singapore) for cyclic amplification. Conditions for amplification were programmed as:

Table 3.3: Conditions for Amplification in a PCR

Steps	Temperature (°C)	Duration (min.)
Initial denaturation	94	4:00
Denaturation	94	1:00
Annealing	57	0:30
Extension	72	1:00
Final extension	72	7:00

The PCR was carried out for a total of 35 cycles.

PCR amplification products were electrophoretically separated on 1.5% agarose gel prepared in TAE (Tris Acetate) buffer (80V) for 1 hour and visualized on gel documentation system (Gel Doc™ XR- BIORAD). DNA fragments were purified using PCR purifying kit. The amplified fungal DNA was submitted for sequencing to Applied Biosystems ABI 3730XL) at the Xcelris Genomics (Ahmedabad, India). Obtained sequences were submitted at US National Centre for Biotechnology Information (NCBI) database for BLAST matching.

3.4. Antibacterial activity of selected mushrooms extracts against two human pathogenic bacteria *Staphylococcus aureus* and *Escherichia coli*.

3.4.1 Materials

Materials used to check antibacterial activity of the fungus were, fruiting bodies of the mushrooms and two bacterial pathogens i.e. *Escherichia coli* and *Staphylococcus aureus*.

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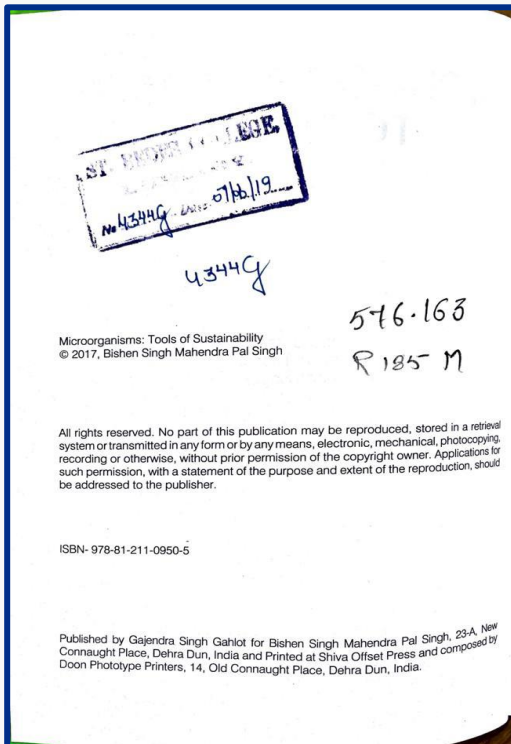
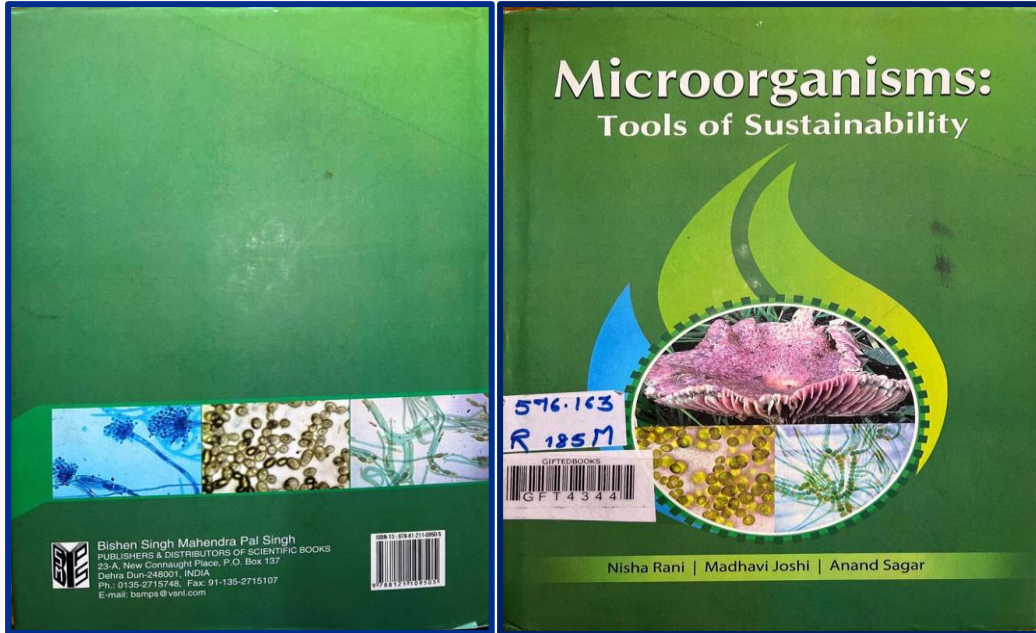
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2. Dr. Madhavi Joshi- Department of Botany
 Title of Book: Microorganisms: Tool of Sustainability
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P. fluorescens, *P. solanacearum*, *P. syringae*, *Serratia entomophila*, *Streptomyces griseoviridis*, *S. lydicus* and various *Rhizobia* spp. However, PGPB inoculated crops represent only a small fraction of current worldwide agricultural practice.

4.2. Plant growth promoting fungi (PGPF)

While most studies have focused on the interaction between rhizosphere bacteria and plant roots, little is known about the plant growth promoting fungi (PGPF) and molecular mechanisms of response of resistance offered by PGPF. The beneficial effects of certain rhizosphere fungi in terms of plant growth promotion and biological control has been reported by many researchers (Windham *et al.* 1986; Hall, 1987; Baker, 1991; Narita and Suzuki, 1991). *Trichoderma harzianum*, *Piriformospora indica*, yeasts like *Saccharomyces cerevisiae*, *Rhodotorula mucilagina* are the most common PGPF (Boby *et al.* 2008). PGPF are nonpathogenic saprophytes and are reported to suppress fungal and bacterial diseases in a number of crop plants (Shivanna *et al.* 1996; Koike *et al.* 2001; Chandanie *et al.* 2006). Fungi have the advantage over bacterial inoculants in that they are generally more effective at spreading through the soil and rhizosphere. Colonization of roots with PGPF can also lead to systemic resistance in distal parts of the plant (Meera, 1995; Munoz *et al.* 2008). Only a few studies of signaling pathways during PGPF-mediated induced systemic resistance, using *Trichoderma* sp. have been performed (Shores, 2005). The PGPF *Phoma* sp., which generally does not sporulate under natural conditions, has been found to improve plant growth, suppress plant pathogens and induce systemic resistance (Hyakumachi and Kubota, 2004). The mechanisms involved in plant growth promotion by fungi are competition with fungal pathogens, antibiotic production and elicitation of defense responses. In addition, many plant beneficial fungi are able to parasitize spores, sclerotia or hyphae of pathogenic fungi, resulting in biocontrol. *Trichoderma* species belong to a class of free living fungi beneficial to plants that are common in the rhizosphere. In addition to their mycoparasitic capabilities, many *Trichoderma* strains are able to colonize and grow in association with plant roots and significantly increase plant growth and development. The beneficial effects of *Trichoderma* depend on more direct mechanisms as certain species including *T. virens* and *T. atroviride* can produce indole3acetic acid (IAA) and other auxin related compounds (Harman *et al.* 2004; Contreras *et al.* 2009).

4.3. Plant Growth Promoting Algae

Since investigations on the ecology of soil algae have shown that this microbial group constitutes an important component of the soil biota, the evaluation of the role played by microalgae in soil economy and plant growth has caused a lively

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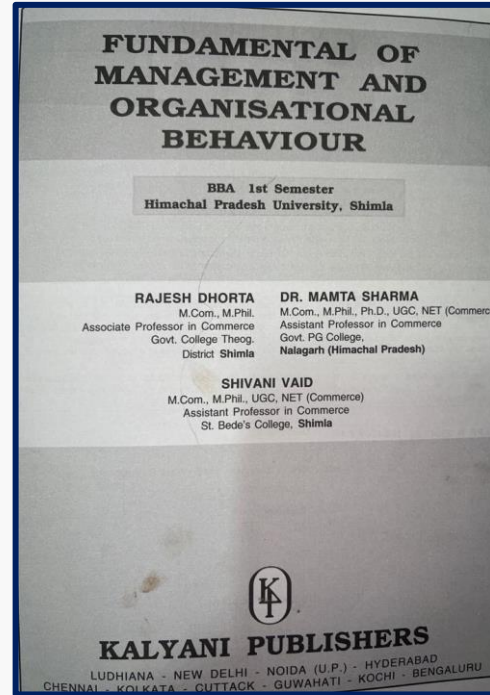
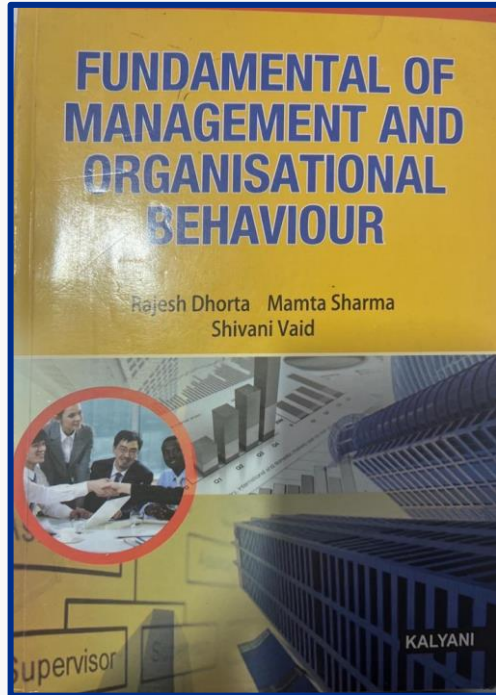
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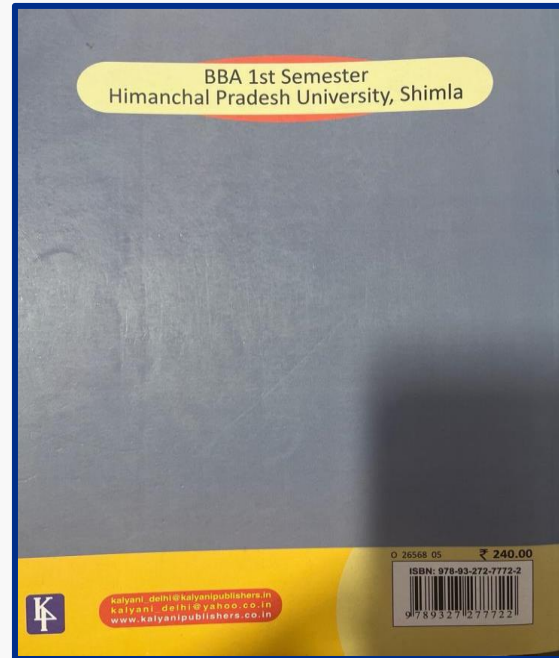
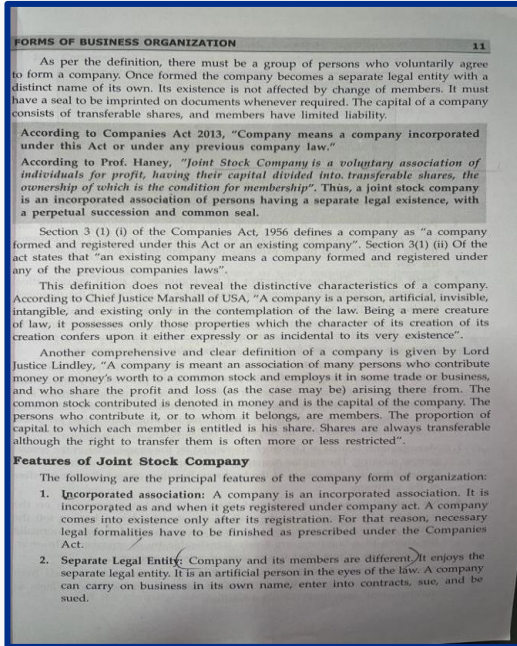


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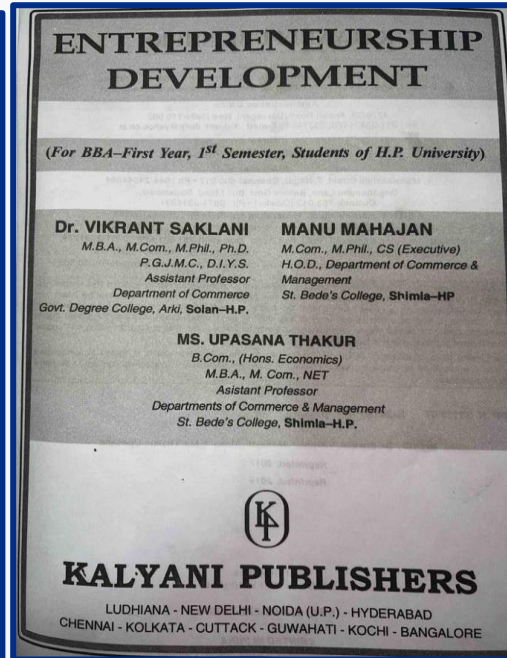
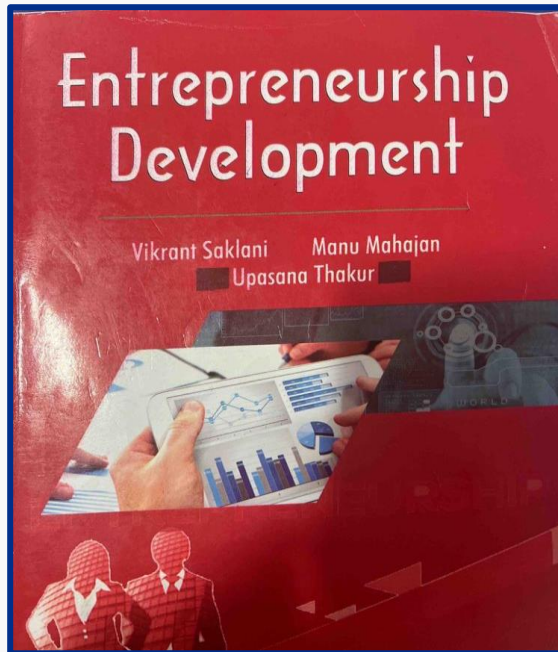
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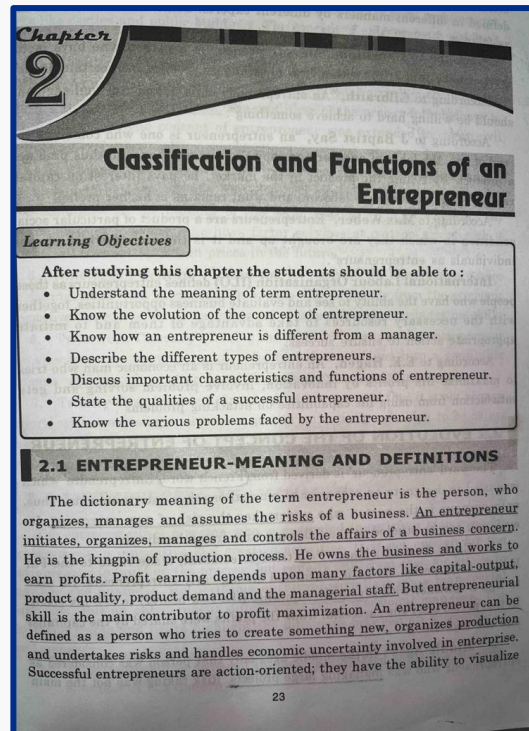
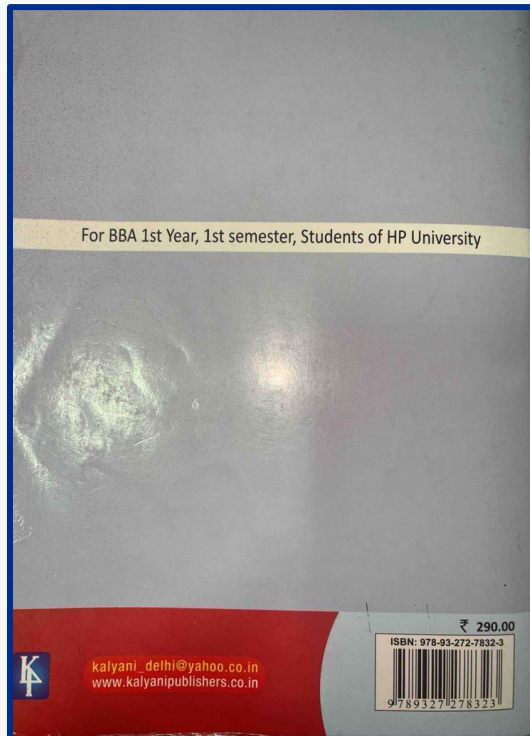
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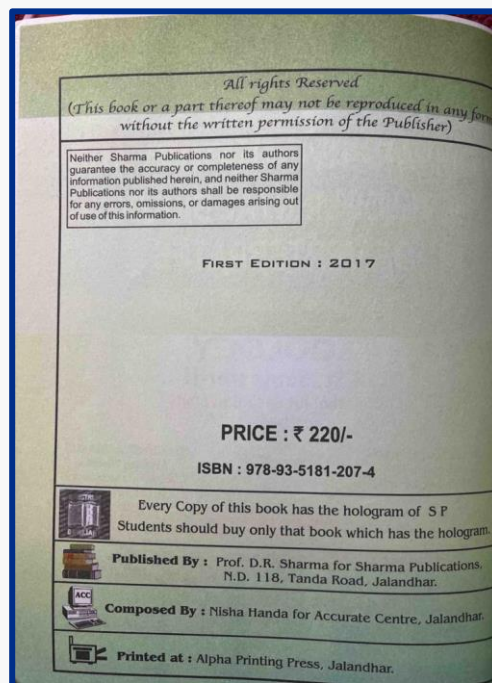
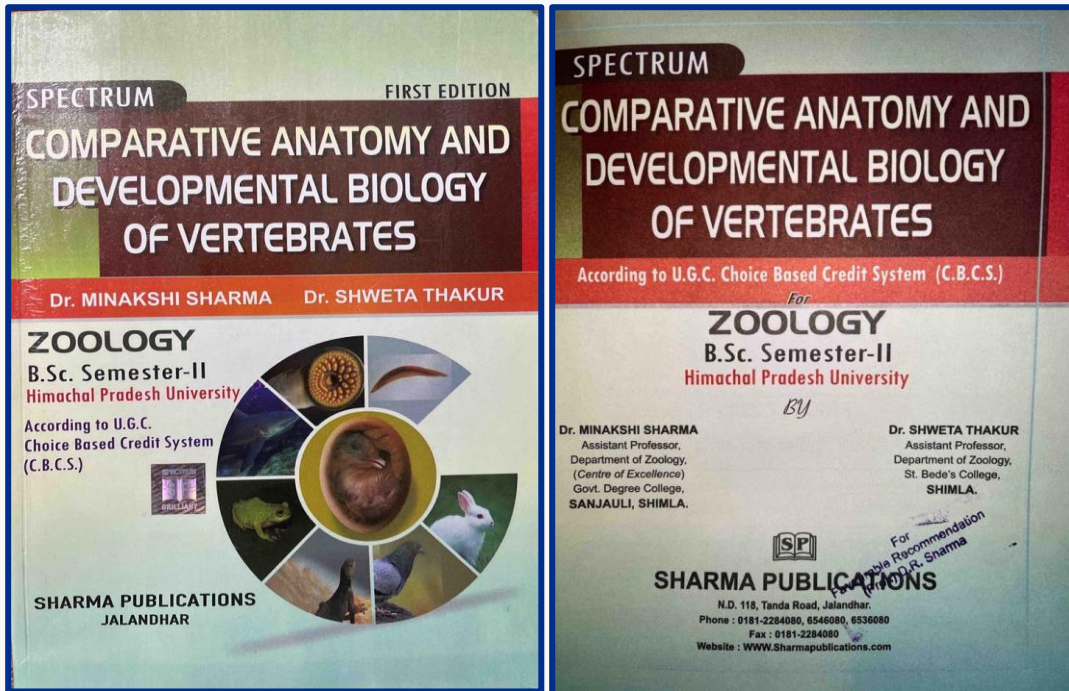
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5. Dr. Shweta Thakur - Department of Zoology

Title of Book: Comparative Anatomy and Developmental Biology of Vertebrates

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54 SPECTRUM COMPARATIVE ANATOMY AND DEVELOPMENTAL BIOLOGY

In turtles, tortoise stomach forms curved tubes.
"J" shaped or "U" shaped stomach is found in elasmobranchs, seal, man.

In Polypterus, stomach appears like a blind pouch due to fusion of cardiac stomach.

In crocodiles and some birds stomach is consisting of anterior proventriculus glands and posterior gizzard or ventriculus.

In ruminants (cow) stomach has four chambers or compartments. Of these chambers (rumen, reticulum, omasum) function as reservoirs of food and the last is true stomach and functions in digestion.

Reticulo-rumen (reticulum and rumen)

Reticulum and rumen are often discussed together since each compartment is a low partition. Eighty percent of the capacity of the stomach is related to the rumen contents of the reticulum and rumen intermix freely. The rumen is the main fermentation chamber where billions of microorganisms attack and break down the relatively indigestible fibre of the ruminant's diet.

Omasum

After fermentation in the reticulum and rumen, food passes to the omasum which acts as a filter pump to sort liquid and fine food particles. Coarse fibre particles do not enter the omasum. Also, the omasum may be the site for absorption of water and nitrogen.

Abomasum

The abomasum is the true stomach and the only site on the digestive tract where gastric juices (HCl and the enzymes, pepsin and rennin). Ingested material remains in the abomasum for 2-3 hours.

In monotremes true stomach is absent.

The diagram illustrates the four chambers of a ruminant's stomach. It shows the Small Intestine at the top left, followed by the Rumen, Reticulum, Omasum, and Abomasum. Arrows indicate the flow of food from the Small Intestine into the Rumen, then to the Reticulum, Omasum, and finally the Abomasum. The Reticulum is shown as a large, sac-like structure, and the Omasum is depicted as a series of overlapping, leaf-like folds.

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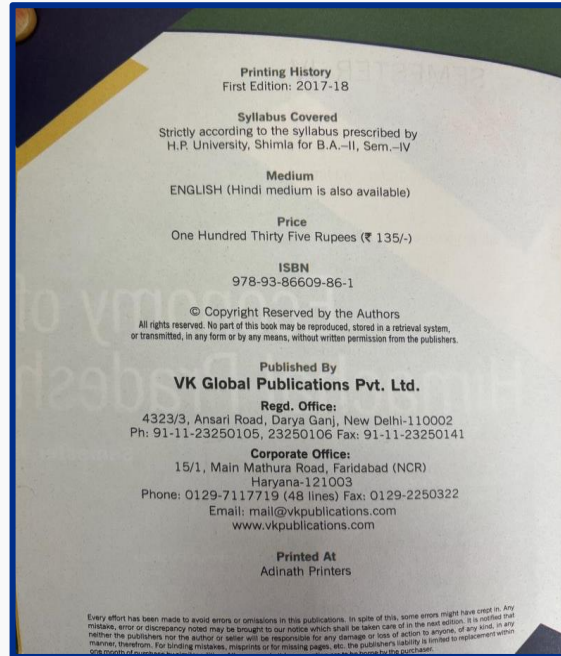
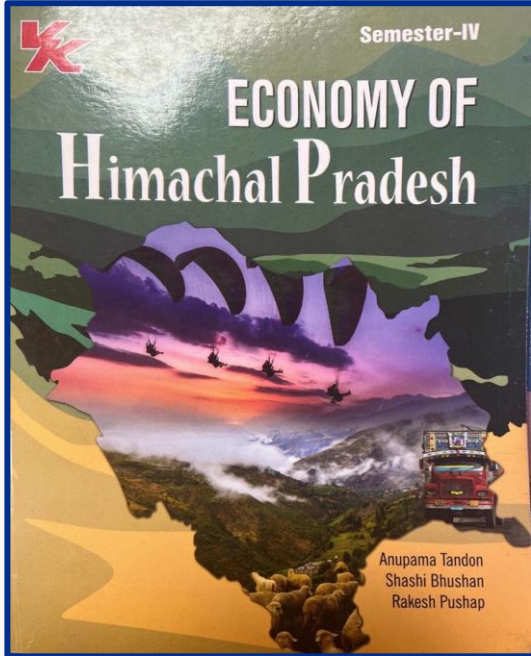
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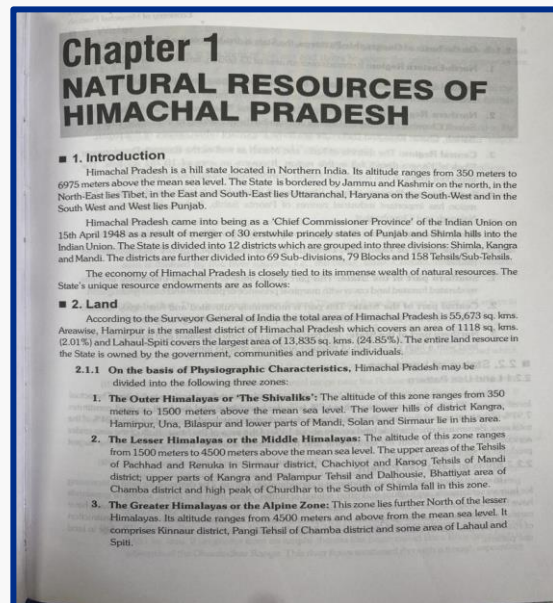
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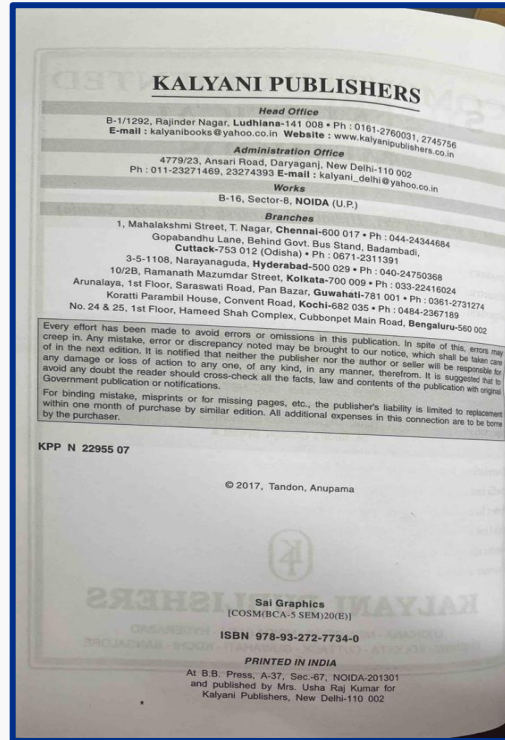
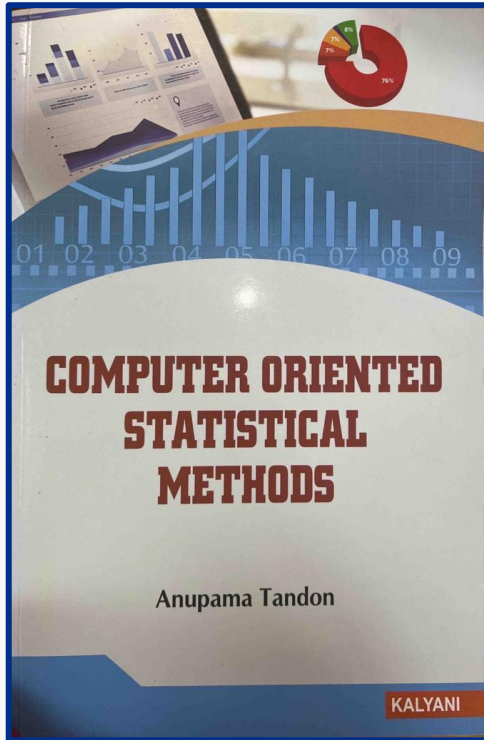
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CLASSIFICATION AND PRESENTATION OF DATA 7

Interval is done by finding the difference between the upper limit of a class and the lower limit of the next higher class and dividing it by 2. The result thus obtained is subtracted from all the upper limits and added to all the upper limits of various classes.

Example 9. Convert inclusive series to exclusive series.

Class Interval

10-19
20-29
30-39
40-49
50-59

$$\text{Adjustment} = \frac{20 - 19}{2} = 0.5$$

Now lower limit of the first class = 10 - 0.5 = 9.5
and upper limit of the first class = 19 + 0.5 = 19.5

Thus following class intervals are obtained after making adjustment in each class.

Class Interval

0.9-19.5
19.5-29.5
29.5-39.5
39.5-49.5
49.5-59.5

1.4 OPEN ENDED DISTRIBUTION

For meaningful comparison between classes, class intervals should be of equal size. In some situations the extreme ends i.e., lower limits and upper limits are not defined. These are called open ended series. In these series mid-value which represents the class cannot be determined and further mathematical calculations cannot be performed. Thus the limits of the open ended class intervals are determined on the basis of other class intervals in the series. But this is possible if the magnitudes of other class intervals are equal.

Example 10.

Marks	No. of Students
Below 30	10
30-40	18
40-50	27
50-60	12
60 and above	8
Total	75

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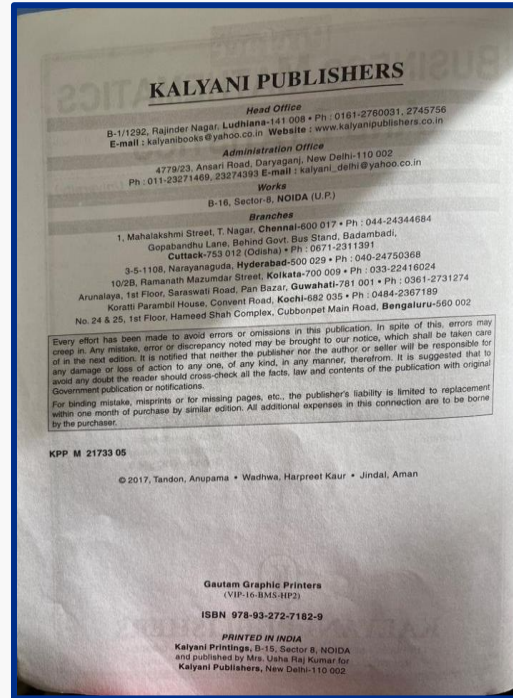
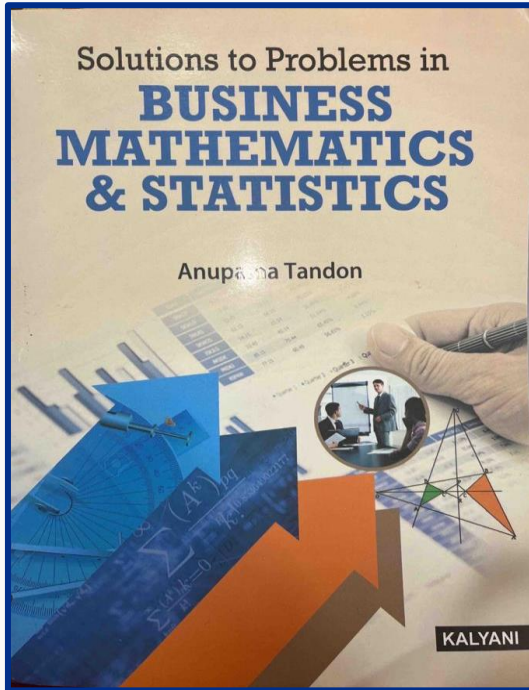
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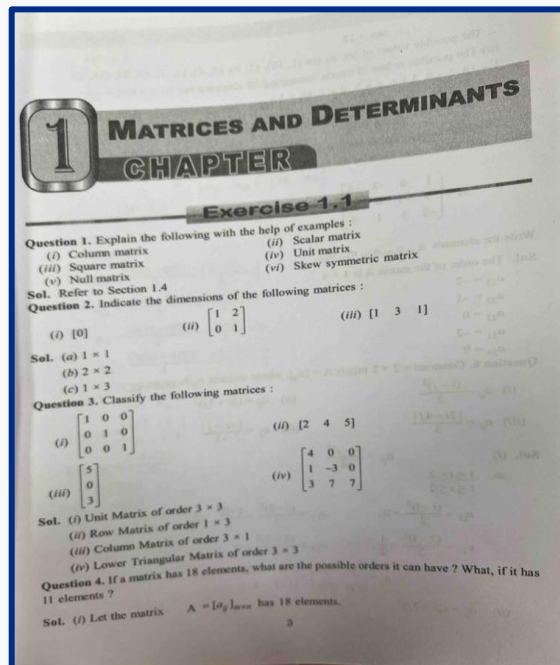
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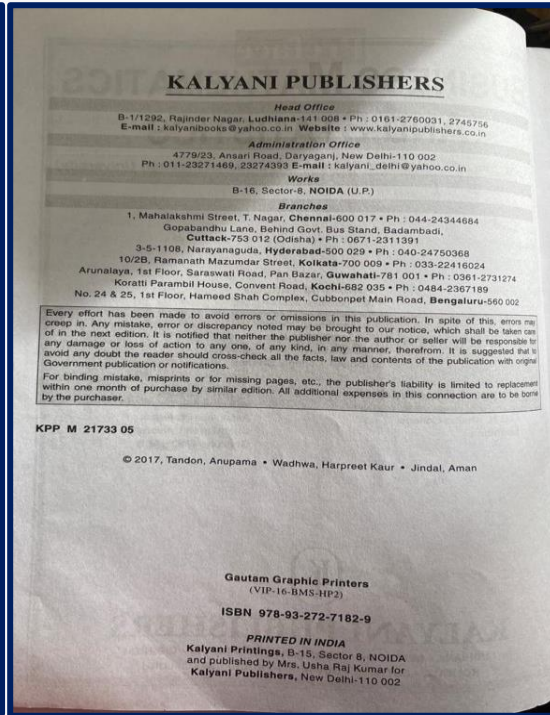
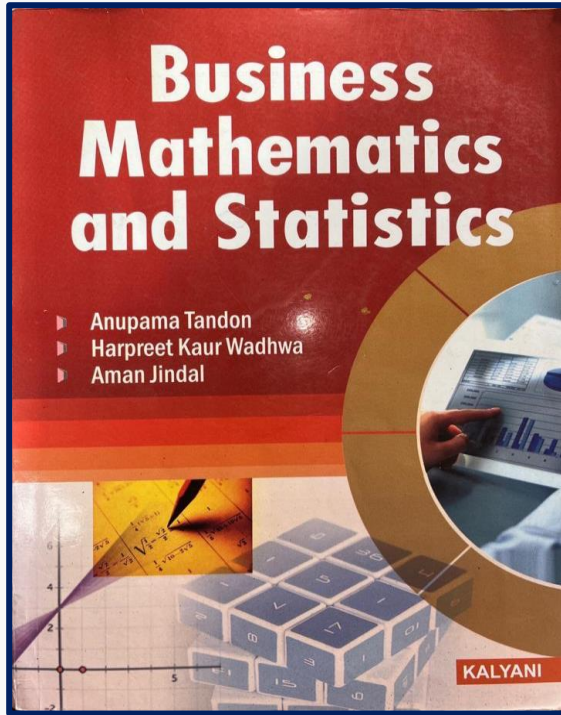
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MATRICES AND DETERMINANTS VI.17

1.8 SCALAR MULTIPLICATION

The matrix obtained by multiplying every element of a matrix A by a scalar k is called the scalar multiple of A by k and is denoted by kA or kA .

If $A = [a_{ij}]$ be an $m \times n$ matrix and k be any complex number, then kA is defined to be the $m \times n$ matrix whose (i, j) th entry is $k a_{ij}$.

For example,

$$A = \begin{bmatrix} a_{11} & a_{12} & a_{13} \\ a_{21} & a_{22} & a_{23} \\ a_{31} & a_{32} & a_{33} \end{bmatrix}$$

If

$$kA = \begin{bmatrix} ka_{11} & ka_{12} & ka_{13} \\ ka_{21} & ka_{22} & ka_{23} \\ ka_{31} & ka_{32} & ka_{33} \end{bmatrix}$$

then

Example 1: If $A = \begin{bmatrix} 6 & 2 & 1 \\ -3 & 9 & 11 \end{bmatrix}$, then find (i) $3A$ (ii) $-2A$ (iii) $0A$.

SOLUTION:

$$A = \begin{bmatrix} 6 & 2 & 1 \\ -3 & 9 & 11 \end{bmatrix}$$

(i) $3A = 3 \begin{bmatrix} 6 & 2 & 1 \\ -3 & 9 & 11 \end{bmatrix} = \begin{bmatrix} 3 \cdot 6 & 3 \cdot 2 & 3 \cdot 1 \\ 3 \cdot (-3) & 3 \cdot 9 & 3 \cdot 11 \end{bmatrix} = \begin{bmatrix} 18 & 6 & 3 \\ -9 & 27 & 33 \end{bmatrix}$

(ii) $-2A = -2 \begin{bmatrix} 6 & 2 & 1 \\ -3 & 9 & 11 \end{bmatrix} = \begin{bmatrix} -2 \cdot 6 & -2 \cdot 2 & -2 \cdot 1 \\ -2 \cdot (-3) & -2 \cdot 9 & -2 \cdot 11 \end{bmatrix} = \begin{bmatrix} -12 & -4 & -2 \\ 6 & -18 & -22 \end{bmatrix}$

(iii) $0A = 0 \begin{bmatrix} 6 & 2 & 1 \\ -3 & 9 & 11 \end{bmatrix} = \begin{bmatrix} 0 \cdot 6 & 0 \cdot 2 & 0 \cdot 1 \\ 0 \cdot (-3) & 0 \cdot 9 & 0 \cdot 11 \end{bmatrix} = \begin{bmatrix} 0 & 0 & 0 \\ 0 & 0 & 0 \end{bmatrix} = 0$

1.9 PROPERTIES OF SCALAR MULTIPLICATION

The following properties of scalar multiplication can be easily proved.

1. Distributive Law: If A and B are comparable matrices and k is any complex number, then $k(A + B) = kA + kB$.
i.e. the scalar multiplication of matrices distributes over the addition of matrices.

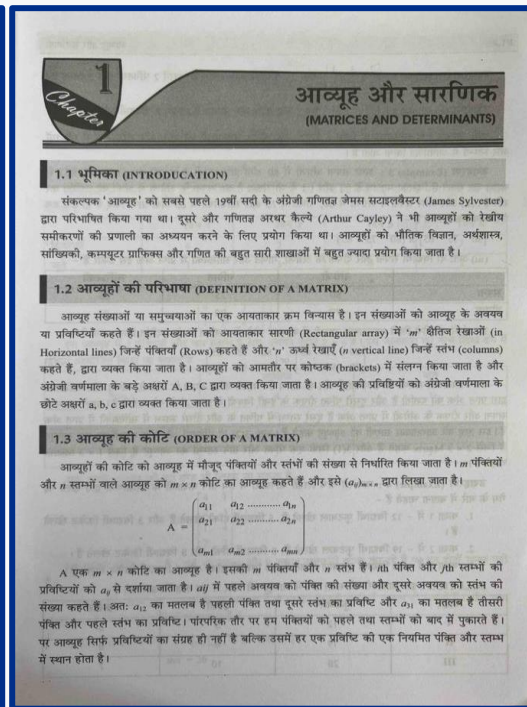
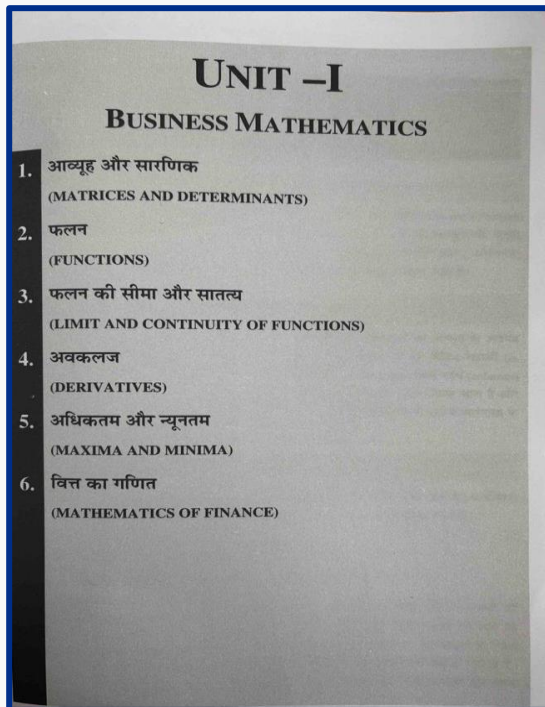
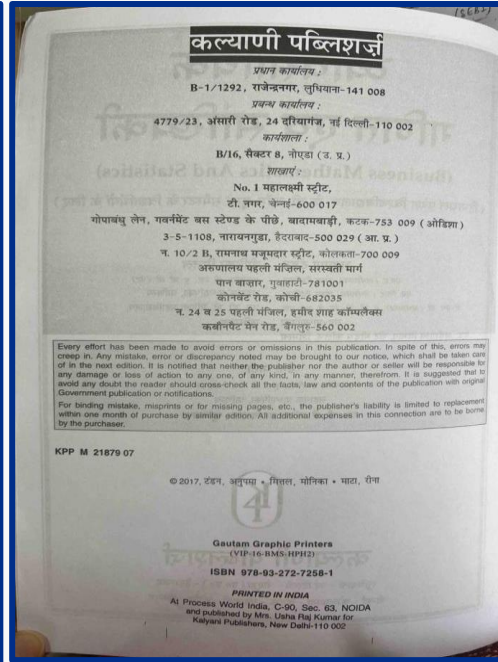
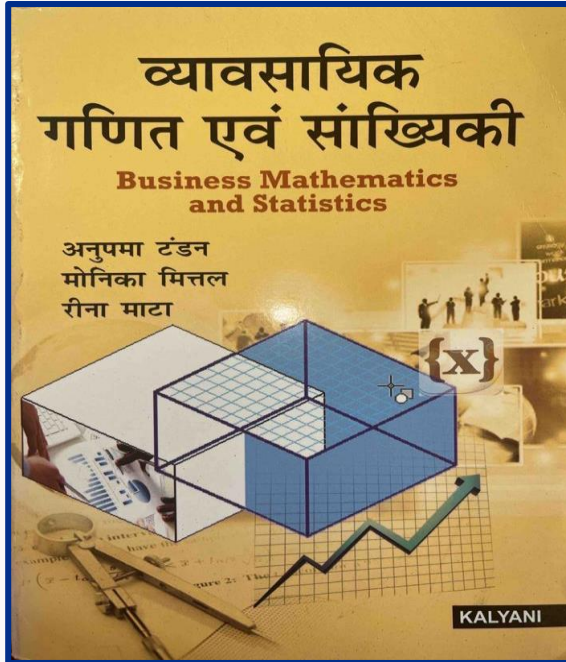
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